

Title:

*Ancient DNA reveals the Arctic origin of Viking Age cod from  
Haithabu, Germany*

Short title:

*Ancient DNA from Viking Age fish*

BIOLOGICAL SCIENCES; GENETICS

SOCIAL SCIENCES; ANTHROPOLOGY

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## **Abstract**

Knowledge of the range and chronology of historic trade and long-distance transport of natural resources is essential for determining the impacts of past human activities on marine environments. However, the specific biological sources of imported fauna are often difficult to identify, in particular if species have a wide spatial distribution and lack clear osteological or isotopic differentiation between populations. Here, we report that ancient fish-bone remains, despite being porous, brittle and light, provide an excellent source of endogenous DNA (15-46%) –of sufficient quality for whole genome reconstruction. By comparing ancient sequence data to that of modern specimens, we determine the biological origin of multiple Viking Age (800-1066 CE) and subsequent medieval (1066-1280 CE) Atlantic cod (*Gadus morhua*) bones from excavation sites in Germany, Norway and the United Kingdom. Archaeological context indicates that one of these sites was a fishing settlement for the procurement of local catches, whereas the other localities were centers of trade. Fish from the trade sites show a mixed ancestry and are statistically differentiated from local fish populations. Moreover, Viking Age samples from Haithabu, Germany are traced back to the North East Arctic Atlantic cod population that has supported the Lofoten fisheries of Norway for centuries. Our results resolve a long-standing controversial hypothesis and indicate that the marine resources of the North Atlantic Ocean were used to sustain an international demand for protein as far back as the Viking Age.

## **Significance**

A rich archaeological record of fish-bone remains testifies to the millennia-long human exploitation of the natural resources of the oceans. In Europe, historical evidence demonstrates that an extensive international industry developed during the Middle Ages that exported preserved cod from the Lofoten Archipelago, northern Norway, to expanding urban centres around the North and Baltic Sea regions. The early origins of this iconic exchange, however, have long been debated. We genetically trace the ancestry of Viking Age fish from mainland Europe to the North East Arctic cod population that supports the modern Lofoten fisheries. This application of genome-wide analyses from ancient fish-bone reveals an early origin of what became an economically important trade, with implications for archaeology and environmental history.

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## **Introduction**

The global trade of animal products is driven by an increasingly international economy that geographically separates consumer demands from their ecological footprint elsewhere (1). Yet long-range trade has an extensive history (2). For instance, in medieval and post-medieval Europe, the onset of urbanized market economies has been linked to the growth of long-range trade by historical and archaeological evidence (3, 4). The exploitation of increasingly distant fish populations has proven to be one of the clearest demonstrations of this ecological globalization (5-9). Fisheries around the coastal regions of the Lofoten Archipelago in Norway have a particularly long history; in this region, the regular arrival of seasonal spawning aggregations of Atlantic cod –migrating southwards from the Arctic Barents Sea (10, 11)– coincides with those climatic conditions essential for the freeze drying and long-term preservation of cod without the use of expensive salt. These unique conditions allowed the development of an extensive long-distance trade and fishery, that for centuries provided a high-quality yet affordable source of protein with a long-shelf-life to urban centers around southern North and Baltic Sea regions (12). Identifying the emergence and growth of such extensive fisheries is a fundamental step in the study of past impacts of human exploitation (13-16). Yet in the centuries before systematic historical records, this crucially depends on the ability to determine the biological source of archaeological samples.

Ancient DNA (aDNA) methods provide unique opportunities to elucidate diversification events, genetic admixture or migration routes (e.g. see 17) and have been used to assign historic individuals to their most likely geographic origin (18-20). In particular, the genome-wide analysis of 1000s or more Single Nucleotide

Polymorphisms (SNPs) allows the determination of an individual's geographic origin with accuracy and precision, even when limited genetic differentiation exists among regions (20-22). Since low levels of genetic differentiation are typical for marine species with high dispersal capabilities like Atlantic cod (23, 24), such genome-wide approaches are essential to increase the assignment power of modern (22) and ancient samples of these species (25). So far, however, aDNA studies employing genome-wide approaches are lacking for marine fishes.

Here, we compare whole genome data of 15 Viking Age and medieval Atlantic cod from five archaeological sites to those of 168 modern specimens from six populations (Fig. 1). We aim to discover where the cod were caught by analyzing fish bones from these sites. For example, were the cod eaten in a Viking Age (800-1066 CE) town of the western Baltic (Haithabu) caught in local waters, in the nearby North Sea or in truly distant waters of Arctic Norway (12), the Northern Isles of Scotland (26) and/or Iceland (6)? All of these distant regions are known to have produced dried fish for export later in the Middle Ages (27).

To improve our ability to identify the source of these ancient samples, we exploit our knowledge of patterns of genomic variation found amongst modern Atlantic cod populations. Distinct genomic regions with elevated population differentiation have been identified for this species (22, 28-30). Several of these regions are co-localized into four mega-base scale, polymorphic inversions with high linkage disequilibrium (31-34). These polymorphisms segregate with a distinct geographical distribution and have been associated with temperature clines and ecotype-specific migratory behavior (31, 32, 34). Their divergence can be used to improve the traceability of individuals (22) and, given their large size (between ~5 and ~17 Mbp), the inversion state of ancient samples can be easily determined from

low coverage sequencing data.

We specifically focus on fish bones from Haithabu (n = 5, dated 800-1066 CE) for several reasons. First, a unique 9<sup>th</sup>-century account records the voyage of a Viking chieftain and trader from Arctic Norway to Haithabu. While his cargo is not fully specified –except for walrus tusks to be gifted to Alfred the Great of England (35)– a mundane consignment of dried cod may not have merited historical record. Second, artefactual evidence has confirmed contact between Haithabu and Scandinavian settlements of the North Atlantic (36). Third, Haithabu (and neighboring Schleswig, which later replaced Haithabu) preceded Lübeck as the main focus of trade in the western Baltic region. Because Lübeck came to control much of Europe's dried cod trade (37) it is possible that its predecessors were already engaged in this exchange –at dates for which the historical record is incomplete. Fourth, the cod finds from Haithabu occurred alongside species such as saithe (*Pollachius virens*), ling (*Molva molva*) and halibut (*Hippoglossus hippoglossus*), which is more consistent with fishing in the North Sea or North Atlantic than in the Kattegat or Baltic Sea (38-40). Finally, isotope analyses using bone collagen (8, 41) or bone carbonate (9) suggest that the Haithabu cod were not locally caught, although their origin remains ambiguous. This previous research pinpoints the fish bones from Haithabu as ideal material with which to test the hypothesis of Viking Age transport of cod from northern Norway using genomic methods. Without definitive interpretation of the Haithabu cod, the earliest secure historical and archaeological evidence for dried cod imports to towns around the North and Baltic Seas dates to the 12<sup>th</sup>, 13<sup>th</sup> and 14<sup>th</sup> centuries (7, 8, 37, 42, 43). Fishing in Arctic Norway occurred long before the Viking Age (44), but most arguments for pre-12<sup>th</sup>-century long-range exports have had to rely on historical sources such as Icelandic sagas that postdate the

events they nominally describe (43, 45).

In addition to the specimens from Haithabu, we include two bones (dated 1100-1280 CE) from Schleswig, one bone (dated 700-950 CE) from the small inland trading settlement of Bjørkum in western Norway and two bones (dated 1025-1175 CE) from Oslo. Lastly, five cod bones (dated 1000-1200 CE) are from a fishing settlement in Orkney, northern Scotland, where a local catch can be confidently assumed (26).

## Results

We obtained 1,013 million paired reads (average read length 40 to 78 bp) that contained 15 to 46% endogenous DNA and resulted in 1- to 3.4-fold nuclear coverage for 15 out of 19 ancient cod samples (Table S1). Analyses of post-mortem degradation patterns showed the typical fragmentation and elevated deamination rates as expected from authentic ancient DNA (Fig. S1). For the modern data (Fig. 1), an average of 47 million paired reads were obtained per specimen, resulting in ~ 9-fold individual coverage of the nuclear genome (Table S2). After SNP calling and filtering we obtained a dataset of 156695 SNPs.

First, we compared the genome-wide diversity of ancient samples and modern samples using principle component (PCA) and ADMIXTURE analyses. We excluded the four chromosomes (LG01, 02, 07, and 12) that contain the large inversions (31-33), as these distort genetic analyses that assume linkage equilibrium. The PCA shows that three modern populations –North East Arctic (NEA), Lofoten and North Sea– form a large, overlapping group, with Iceland clustering near the NEA (Fig. 2a). The majority of the ancient samples cluster with this large group. The modern eastern Baltic and Øresund form two distinct groups, with one ancient specimen from Schleswig clustering with the Øresund. A lack of strong genetic population structure between most modern samples is supported by ADMIXTURE, for which the best fit (based on lowest CV-error) is two populations ( $k = 2$ ). Here, Iceland and eastern Baltic have distinct ancestry, whereas the other modern populations show a more mixed ancestry (Fig. 2b). The Øresund has a higher level of Baltic ancestry than the other admixed populations. All ancient individuals have mixed ancestry, with the one Schleswig sample having a similar level of Baltic ancestry as the modern Øresund. At a lower optimal fit, ADMIXTURE estimates biologically plausible ancestry for  $k = 3$



(identifying an Øresund component) and  $k = 4$  (identifying a North Sea component). Nonetheless, for  $k = 5$  the model separates the original eastern Baltic cluster rather than differentiating either NEA or Lofoten cod (Fig. S2). Overall, these analyses show that the ancient individuals are genetically most related to four modern populations; one Schleswig individual is genetically more similar to the Øresund and all other individuals clearly classify to the NEA, Lofoten, or North Sea population.

To further differentiate the origins of the ancient individuals, we determined their genotypes for the inversion loci on LG01, 02, 07 and 12. Modern populations can have divergent allele frequencies for these loci (31, 32) and the NEA population is set apart by having inverted alleles near fixation at all four loci (Fig. 3a). PCAs of these inversions for the four modern populations show the typical tri-modal clustering of a bi-allelic locus with heterozygote genotypes clustering intermediate to the two divergent homozygotes (46) (Fig. S3). All ancient individuals cluster within this tri-model pattern, allowing for the decisive determination of their genotypes at each inversion locus (Fig. S3). Collinear alleles dominate in the Orkney samples, while inverted alleles dominate in Haithabu (Fig. 3b). The Bjørkum sample has a predominantly inverted composite genotype, whereas Schleswig and Oslo show a mixed pattern, with one specimen having a collinear and the other having a more inverted composite genotype, in each location respectively.

Based on their inversion genotypes we investigated if ancient individuals statistically differed in their resemblance towards a particular modern population. The probability of obtaining an inversion genotype follows a binomial distribution given the underlying allele frequency in a population. We can assume independence between loci since the inversions are located on different chromosomes. It is thus straightforward to calculate the overall probability of obtaining a composite ancient

inversion genotype –based on the four modern populations’ respective allele frequencies – as a measure of an individual’s affinity towards a specific population. Based on this probability, we find that –apart from a single Haithabu specimen– the ancient samples show two types of affinity (Fig. 3c); individuals with a predominantly collinear composite genotype have a  $> 99\%$  probability of being drawn from either the Lofoten, the North Sea or the Øresund population. Conversely, individuals with a predominantly inverted composite genotype have a  $> 99\%$  probability of being drawn solely from the NEA population. The atypical Haithabu specimen has a  $> 99\%$  probability of coming from either the NEA or Lofoten population.

## Discussion

By investigating genome-wide patterns of variation –including four megabase-scale inversions (31-34)– we show that most ancient cod specimens from Viking Age (Haithabu and Bjørkum) and early medieval (Schleswig and Oslo) trading sites have a near exclusive genomic affinity to the modern North East Arctic population. This finding has archaeological and evolutionary implications.

First, this study provides a unique genome-wide study of archaeological fish bone and demonstrates its potential as an archive of ancient DNA. We obtained short reads (< 100 bp), with the typical fragmentation and cysteine de-amination patterns expected after post-mortem degradation (47-49). These results are therefore fully consistent with the extraction and analysis of authentic ancient Atlantic cod DNA. Not all types of animal bone preserve DNA equally well and significantly higher proportions of endogenous DNA have been found in dense and heavy bone types such as the petrous bone (50-52). It is presumed that it is the high density of the bone that leads to reduced bacterial and chemical-mediated decay and improved DNA preservation (50, 53). This hypothesis would suggest that fish bones –that are porous, brittle and light– should be a poor source of DNA. Instead, we find surprisingly high levels (15-46%) of endogenous DNA preservation in 15 out of 19 fish bone specimens up to 1300 years old from five different archeological sites. Notwithstanding the use of a novel extraction protocol aimed to maximize endogenous DNA (54), this rate of success compares favorably to results from mammalian bones whereby the majority of samples –excluding petrous bones– typically yield a few percent endogenous DNA at most (55). Our positive results agree with studies using PCR-based methods that have reported successful amplification from fish bones (56-60) in some cases up to 10,000 years old (61). The

observation that porous, light fish bones can yield whole genome shotgun libraries with high levels of endogenous DNA underscores our lack of understanding of DNA preservation in different types of animal bone. These results also illuminate the potential of the large reservoir of archaeological fish bone as a source for aDNA of sufficient quantity and quality to study long-term evolutionary processes in the marine environment.

Second, we identify polymorphic chromosomal inversions in ancient Atlantic cod specimens. Chromosomal inversions are expected to play a major role in ecological adaptation (62, 63). In cod, these regions contribute to elevated genomic diversification between modern populations and ecotypes –despite low overall levels of divergence (25, 31-34, 64)– and they have been suspected to be under selection (22, 28, 29). The high genomic divergence of these alleles, in combination with their wide geographic distribution, suggests that these have been maintained as a polymorphism for hundreds of thousands, if not millions of years (32, 33). Here, we directly observe the millennium-long maintenance of such inversions, which provides an opportunity to investigate their temporal stability. Although speculative due to the currently small sample sizes from Orkney ( $n = 5$ ) and Haithabu ( $n = 5$ ), our results suggest that the divergent frequency distributions of these inversions indeed have remained stable in their respective populations. Given the low overall genomic differentiation outside of these inversions in modern populations (31-33), it appears that this divergence is sustained despite ongoing gene flow, which further supports a hypothesis that these alleles are maintained as a balanced polymorphism (65).

Finally, we solve a long-standing hypothesis generated by history, archaeology, and stable isotope analysis that dried cod from northern Norway were transported during the Viking Age to Haithabu. Currently, the NEA cod feeds in the

Barents Sea and its spawning grounds are restricted to the northern coasts of Norway, especially (although not exclusively) in the area around Lofoten (10, 11). One could argue that this population spawned in the Skagerrak or Kattegat during the Viking Age, but such a scenario is implausible; historical records of the Norwegian fisheries since the 12<sup>th</sup> century show that spatial fluctuations in the distribution of fishing effort targeting these spawning aggregations have been restricted (43, 66). Furthermore, our ancient data are also consistent with observed long-term spawning fidelity in the North Sea region (67, 68); the genetic affinity of the ancient Orkney population, a focus of fishing since the Viking Age (69), agrees with those of modern individuals living in the North Sea. Similarly, we observe the specific affinity of one ancient Schleswig specimen towards a distinct population in the Øresund (70). This observation also suggests local spawning fidelity and agrees with fine-scale population structure earlier found in this region (70-73). Overall, by identifying their most likely source population, we conclude that the ancient Haithabu cod were not caught locally, but around the Norwegian coast during historic spawning aggregations of NEA cod. Since climatic conditions restrict the production of dried cod without salt to the north of Norway (66), we can further constrain their source to this northern region, implying transportation over large spatial distances during the Viking Age. We cannot yet infer conclusions about the scale of this activity, potentially ranging from providing travelers' rations to supplying an urban staple, which –together with obtaining a more refined chronology within the Viking Age– will be a subject of future research. Moreover, knowing the Haithabu results, it becomes important to ask whether cod bones from even earlier trading sites in the western Baltic region (e.g. Groß Strömkendorf) might also represent fish from a distant source (74).

## **Conclusion**

Our discovery of distinctive genomic inversions in ancient cod specimens has made it possible to answer the long-standing question of whether dried cod was transported from northern Norway during the Viking Age (800-1066 CE), solving a mystery epitomized by the 9<sup>th</sup>-century account of an Arctic Norwegian chieftain's voyage to Haithabu. Our findings suggest that distant requirements for Arctic protein had thus already begun to influence the economy and ecology of the north over the chronology under consideration. Our study highlights the potential of coupling modern genomics with ancient DNA to study the origins of historic trade routes in fish and other taxa.

## Material and Methods

The ancient samples ( $n = 19$ ) are from the archives of excavations conducted at Haithabu (39), Schleswig (75), Bjørkum (76), Oslo (77) and Orkney (26) and are dated based on archaeological context. They come from waterlogged (Haithabu, Schleswig and Oslo) and free-draining (Bjørkum and Orkney) deposits and have been stored dry and unfrozen in fluctuating ambient temperatures after excavation. Bones were morphologically identified as Atlantic cod and selected from different archaeological layers, from fish of differing size and/or from the same element to avoid multiple samples from individual fish (Fig. S4).

**Extraction and library creation:** DNA from ancient samples was extracted in a dedicated aDNA laboratory at the University of Oslo following strict precautions (78, 79) using a combined bleach and pre-digestion (BleDD<sup>2</sup>) protocol (54). Ancient DNA libraries were created using a blunt-end ligation protocol (80) with minor adjustments (20) (Supplementary Information). DNA from modern samples ( $n = 168$ , Fig. 1, Table S2) was extracted—in a separate laboratory from the ancient samples—using a DNeasy Blood & Tissue kit (Qiagen), and sheared to an approximate insert size of 350bp (81). Modern libraries were created using a TruSeq DNA PCR-Free Preparation Kit. All libraries were sequenced on an Illumina HiSeq 2500.

**Data processing:** The ancient read data were processed using PALEOMIX (82). In short, forward and reverse reads were collapsed with AdapterRemoval v1.5 (83) and aligned to the Gadmor2 reference (84, 85) using BWA *aln* v.0.7.5a-r405 (86). The modern data were aligned using BWA *mem*. Reads that aligned with a minimum quality score (MapQ) of 25 used for subsequent analyses. aDNA damage patterns were investigated using mapDamage v.2.0.6 (49).

SNP genotypes were obtained using GATK v. 3.4.46 (87), after duplicate

removal (Picard Tools v. 1.96) and indel realignment (*GATKs IndelRealigner*). Genotypes were jointly (*GATKs Genotypecaller*) called for modern and ancient samples separately with default settings, allowing a maximum of three alternate alleles. The modern data were filtered with BCFTOOLS v. 1.3 (88) using filter `-e "FS>60.0 || MQRankSum<-12.5 || ReadPosRankSum<-8.0 || QD<2.0 || MQ<40" -- SnpGap 10` and VCFTOOLS v.0.1.14 (89), keeping bi-allelic loci with a maximum average read depth of 30 and a minimum MAF of 0.05. The filtered modern and ancient dataset were intersected (BCFTOOLS *isec*), after which genotypes with a quality below 15 and read depth below 3 were set as missing and all C>T and G>A SNPs were removed. The final dataset consisted of 156695 SNPs.

**Analyses:** For inferring genome-wide population structure, SNPS were pruned (`--indep-pairwise 100 10 0.5`) for linkage disequilibrium (LD) using PLINK v1.90p (90) and LG01, 02, 07 and 12 were excluded. PCA was performed with smartPCA, EIGENSOFT v.6.1.4 (91) whereby ancient individuals were "projected" using 'lsqproject' to account for missing data. Model-based clustering was performed using ADMIXTURE v1.3 (92). PCA plots for the inverted regions (LG01; 9.1 - 26.2 Mbp, LG02; 18.5 - 24 Mbp, LG07; 13.6 - 23 Mbp and LG12; 1.3 -13.6 Mbp) were generated without LD pruning (46). Finally, the probability of obtaining the ancient individual's composite inversion genotype from the allele frequency distribution of the four modern populations was calculated by binomial sampling of genotypes and scaling these probabilities to one.

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### **Data availability**

All ancient read data are available at the European Nucleotide Archive (ENA, [www.ebi.ac.uk/ena](http://www.ebi.ac.uk/ena)) under study accession number PRJEB20524 and PRJEB15516. The filtered SNP dataset is available at Dryad (<http://datadryad.org/>) with project number XXXXXX.

### **Author contributions**

BS & JHB designed the study. BS & SB performed the analyses. BS & JHB interpreted the results with input from SB, EN, DH, KSJ, SJ & NCS. ATG & HN did the ancient DNA laboratory work. JHB selected ancient samples provided by EN, AKH, DH & JHB. CP, HK, CA, JD, CP & SJ sampled and contributed the modern samples. KSJ, SJ & NCS managed logistics, provided funding and consumables. BS & JHB wrote the paper with input from all authors.

### **Conflict of Interest**

The authors declare no conflict of interest

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## Figure Legends

**Fig. 1.** Approximate sampling locations of Atlantic cod in the northern Atlantic region in Europe. **a)** Atlantic cod (*Gadus morhua*) specimens (sample size is indicated between brackets) were obtained from modern populations (*black*) and archaeological excavations (*red*). The North East Arctic (NEA) sample was obtained in winter, when this population migrates southwards from the Barents Sea to the Lofoten Archipelago to spawn. The Lofoten population was sampled during summer when the NEA cod are absent from this region. The modern range of Atlantic cod is indicated by blue shading. **b)** Archaeological Atlantic cod jaw-bone (premaxilla) from Orkney.

**Fig. 2.** Genetic population structure in 183 Atlantic cod specimens. **a)** Principle component analysis (PCA) based on 99819 SNPs. Ancient specimens (*stars*) were projected onto the first two principle components calculated using individuals from modern populations (*circles*). **b)** ADMIXTURE ancestry components ( $k = 2$ ) for modern and ancient specimens. The width of the bar for ancient specimens is widened to aid visualization. Linkage group (LG) 01, 02, 07, 12 and unplaced scaffolds were excluded from these analyses (see text for explanation).

**Fig. 3.** Spatial genomic variation in megabase-scale inversions in Atlantic cod. **a)** Allele frequency distribution of four inversions (on LG01, 02, 07 and 12) in four modern populations. The collinear allele (grey) and inverted allele (yellow) segregate as biallelic loci. **b)** Individual inversion genotypes of ancient Atlantic cod. The collinear (grey, AA), inverted (yellow, BB) and heterozygote (yellow/grey, AB) genotypes segregate independently on four chromosomes. **c)** Genotypic affinity of ancient specimens. The overall probability of obtaining the ancient individual's composite genotype was calculated by binomial sampling of inversion genotypes from the respective allele frequency distributions of the four modern populations.



## Figure Legends Supplementary Information

**Fig. S1.** aDNA fragmentation and mis-incorporation patterns of sequencing read data from 15 Atlantic cod samples. Patterns were obtained using MapDamage v. 2.0.6 after down-sampling BAM files to 1,000,000 reads. For visualization purposes, we only show the typical increase in C->T mis-incorporations due to cytosine deamination at the 5'-end of DNA fragments and the corresponding increase of G->A mis-incorporations at the 3'-end.

**Fig. S2.** ADMIXTURE ancestry components for modern and ancient Atlantic cod specimens. Population structure was investigated using models with a variable number of clusters (k). Model fit was assessed by calculating the cross-validation (CV) error, with a lower CV error indicating a better fit. Linkage group (LG) 01, 02, 07, 12 and unplaced scaffolds were excluded from these analyses (see text for explanation).

**Fig. S3.** Principle component analysis of genomic inversions in Atlantic cod. Ancient specimens (*stars*) were projected onto the first two principle components calculated using individuals from modern populations (*circles*). The first principle component (PCA 1) separates genomic variation within each of the four mega-base long regions (LG01; 9.1 - 26.2 Mbp, LG02; 18.5 - 24 Mbp, LG07; 13.6 - 23 Mbp and LG12; 1.3 - 13.6 Mbp) into distinct clusters (*grey dotted ovals*) that reflect the bi-allelic segregation of the three major inversion genotypes (AA; collinear, AB; heterozygote and BB; inverted). The number of SNPs (*n*) used per region is indicated. Mean heterozygosity values per genotype (*presented in grey under each genotype*; estimated by calculating the inbreeding coefficient *F* using a method of moments as implemented in VCFTOOLS v0.1.14) show the marked decrease in *F*-values for the

AB genotypes due to heterozygote excess. Ancient samples follow the tri-modal cluster pattern of the modern individuals.

## Supporting Information:

### *Ancient DNA reveals the Arctic origin of Viking Age cod from Haithabu, Germany*

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## Supplementary Figure Legends

**Fig. S1.** aDNA fragmentation and mis-incorporation patterns of sequencing read data from 15 Atlantic cod samples. Patterns were obtained using MapDamage v. 2.0.6 after down-sampling BAM files to 1,000,000 reads. For visualization purposes, we only show the typical increase in C->T mis-incorporations due to cytosine deamination at the 5'-end of DNA fragments and the corresponding increase of G->A mis-incorporations at the 3'-end.

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## Supplementary Tables

**Table S1.** Specimen ID, location, estimated date and bone type of ancient Atlantic cod samples. All bones were morphologically identified as Atlantic cod (*Gadus morhua*). WGS shotgun libraries were paired-end sequenced, and we report the number of collapsed reads, their clonality, their endogenous DNA content (defined as the unique, non-repetitive fraction of reads aligning towards the gadmor2 reference genome with a minimum MapQ value of 25), the average insert length and the fold coverage obtained for the nuclear genome. Four other specimens (not shown) were extracted of which two did not yield libraries, and two had endogenous DNA content below 1%.

Specimen	Country	Location	Date (CE)	Bone type	Reads (millions)	Clonality (%)	Endogenous DNA (%)	Average insert length (bp)	Fold coverage
COD003	Germany	Schleswig	c.1100-1200	Vertebra	52	14	33	69	1.9
COD023	Germany	Schleswig	c.1200-1280	Articular	67	6	37	51	2.0
COD027	UK	Orkney	c.1000-1200	Premaxilla	59	9	42	75	2.9
COD028	UK	Orkney	c.1000-1200	Premaxilla	84	4	15	72	1.4
COD029	UK	Orkney	c.1000-1200	Premaxilla	58	5	33	71	2.1
COD030	UK	Orkney	c.1000-1200	Premaxilla	55	8	25	72	1.6
COD034	UK	Orkney	c.1000-1200	Premaxilla	49	23	27	72	1.5
COD053	Germany	Haithabu	c.800-1066	Cleithrum	83	8	28	47	1.7
COD054	Germany	Haithabu	c.800-1066	Cleithrum	58	8	37	67	2.3
COD061	Germany	Haithabu	c.800-1066	Vertebra	72	11	24	40	1.1
COD062	Germany	Haithabu	c.800-1066	Vertebra	76	8	25	46	1.4
COD063	Germany	Haithabu	c.800-1066	Articular	95	6	32	46	2.2
COD076	Norway	Bjorkum	c.700-950	Dentary	74	27	18	48	1.0
COD086	Norway	Oslo	c.1025-1175	Ceratohyal	59	12	46	78	3.4
COD092	Norway	Oslo	c.1025-1175	Cleithra	72	16	21	47	1.2

**Table S2.** Population, specimen ID and sample date of modern Atlantic cod specimens. We report if sampling took place during spawning, the number of reads obtained and the resulting fold coverage for the nuclear genome.

Population	Specimen ID	Sample Date	Spawning population	Number of reads (millions)	Fold coverage
Eastern Baltic	ARK_4001	2012, May	Yes	40	7.4
Eastern Baltic	ARK_4002	2012, May	Yes	20	3.8
Eastern Baltic	ARK_4003	2012, May	Yes	38	6.9
Eastern Baltic	ARK_4006	2012, May	Yes	34	6.3
Eastern Baltic	ARK_4007	2012, May	Yes	34	6.4
Eastern Baltic	ARK_4008	2012, May	Yes	47	8.8
Eastern Baltic	ARK_4010	2012, May	Yes	31	5.9
Eastern Baltic	ARK_4011	2012, May	Yes	49	9.1
Eastern Baltic	ARK_4012	2012, May	Yes	35	6.6
Eastern Baltic	ARK_4013	2012, May	Yes	37	6.9
Eastern Baltic	ARK_4014	2012, May	Yes	34	6.1
Eastern Baltic	ARK_4015	2012, May	Yes	44	8.3
Eastern Baltic	ARK_4016	2012, May	Yes	29	5.1
Eastern Baltic	ARK_4017	2012, May	Yes	45	8.5
Eastern Baltic	ARK_4018	2012, May	Yes	33	6.1
Eastern Baltic	ARK_4019	2012, May	Yes	47	8.8
Eastern Baltic	ARK_4020	2012, May	Yes	23	4.1
Eastern Baltic	ARK_4021	2012, May	Yes	57	10.6
Eastern Baltic	ARK_4022	2012, May	Yes	34	6.4
Eastern Baltic	ARK_4024	2012, May	Yes	31	5.8
Eastern Baltic	ARK_4030	2012, May	Yes	34	6.5
Eastern Baltic	ARK_4032	2012, May	Yes	91	17.2
Eastern Baltic	ARK_4039	2012, May	Yes	63	11.9
Eastern Baltic	ARK_4043	2012, May	Yes	68	12.8
Eastern Baltic	BOR_90E	2011, April	Yes	44	8.3
Eastern Baltic	BOR_91E	2011, April	Yes	47	9.0
Eastern Baltic	BOR_74E	2011, May	Yes	45	8.6
Eastern Baltic	BOR_79E	2011, May	Yes	50	9.6
Eastern Baltic	BOR_60E	2012, April	Yes	46	8.7
Eastern Baltic	BOR_611E	2012, May	Yes	45	8.5
Eastern Baltic	BOR_613E	2012, May	Yes	43	8.2
Eastern Baltic	BOR_614E	2012, May	Yes	42	7.9
Eastern Baltic	BOR_615E	2012, May	Yes	42	8.0
Eastern Baltic	BOR_617E	2012, May	Yes	41	7.8
Eastern Baltic	BOR_621E	2012, May	Yes	41	7.8
Eastern Baltic	BOR_624E	2012, May	Yes	42	8.1
Eastern Baltic	BOR_630E	2012, May	Yes	40	7.5
Eastern Baltic	BOR_655E	2012, May	Yes	73	13.8
Eastern Baltic	BOR_659E	2012, May	Yes	59	11.1
Eastern Baltic	BOR_664E	2012, May	Yes	66	12.5
Eastern Baltic	BOR_AL713	2012, May	Yes	44	8.4
Eastern Baltic	BOR_739E	2012, May	Yes	57	10.8
Eastern Baltic	BOR_749E	2012, May	Yes	59	11.2
Eastern Baltic	BOR_760E	2012, May	Yes	6	1.1
Eastern Baltic	BOR_AL724	2012, May	Yes	49	9.3
Eastern Baltic	BOR_AL736	2012, May	Yes	44	8.4
Eastern Baltic	BOR_AL741	2012, May	Yes	46	8.7
Eastern Baltic	BOR_AL777	2012, May	Yes	43	8.3
Iceland	I50_02	2003, April	Yes	49	10.9
Iceland	I50_03	2003, April	Yes	43	9.7
Iceland	I50_04	2003, April	Yes	44	9.8
Iceland	I50_05	2003, April	Yes	46	10.3
Iceland	I50_06	2003, April	Yes	32	7.1
Iceland	I50_07	2003, April	Yes	42	9.4
Iceland	I50_08	2003, April	Yes	35	7.7
Iceland	I50_09	2003, April	Yes	51	11.2
Iceland	I50_10	2003, April	Yes	43	9.6
Iceland	I50_11	2003, April	Yes	50	11.1
Iceland	I50_12	2003, April	Yes	39	8.8
Iceland	I50_13	2003, April	Yes	48	10.7
Iceland	I50_14	2003, April	Yes	26	5.7
Iceland	I50_15	2003, April	Yes	41	9.3
Iceland	I50_16	2003, April	Yes	49	11.0
Iceland	I50_17	2003, April	Yes	43	9.5
Iceland	I50_18	2003, April	Yes	46	10.1

Iceland	I50_19	2003, April	Yes	52	11.4
Iceland	I50_20	2003, April	Yes	54	12.1
Iceland	I50_23	2003, April	Yes	58	12.9
Iceland	I50_26	2003, April	Yes	53	10.3
Iceland	I50_38	2003, April	Yes	64	12.3
Iceland	I50_41	2003, April	Yes	92	19.1
Iceland	I50_42	2003, April	Yes	52	11.6
Lofoten	LOF_A_14_01	2014, August	No	60	11.4
Lofoten	LOF_A_14_03	2014, August	No	54	10.2
Lofoten	LOF_A_14_04	2014, August	No	47	9.0
Lofoten	LOF_A_14_05	2014, August	No	47	8.9
Lofoten	LOF_A_14_06	2014, August	No	52	10.0
Lofoten	LOF_A_14_08	2014, August	No	43	8.2
Lofoten	LOF_A_14_09	2014, August	No	50	9.5
Lofoten	LOF_A_14_10	2014, August	No	48	9.1
Lofoten	LOF_A_14_11	2014, August	No	53	10.1
Lofoten	LOF_A_14_16	2014, August	No	54	10.1
Lofoten	LOF_A_14_17	2014, August	No	60	11.4
Lofoten	LOF_A_14_18	2014, August	No	52	9.9
Lofoten	LOF_A_14_19	2014, August	No	54	10.3
Lofoten	LOF_A_14_20	2014, August	No	43	8.2
Lofoten	LOF_A_14_21	2014, August	No	52	9.8
Lofoten	LOF_A_14_22	2014, August	No	48	9.1
Lofoten	LOF_A_14_23	2014, August	No	46	8.8
Lofoten	LOF_A_14_24	2014, August	No	49	9.3
Lofoten	LOF_A_14_25	2014, August	No	59	11.2
Lofoten	LOF_A_14_26	2014, August	No	51	9.7
Lofoten	LOF_A_14_27	2014, August	No	65	12.4
Lofoten	LOF_A_14_28	2014, August	No	49	9.3
Lofoten	LOF_A_14_29	2014, August	No	46	8.7
Lofoten	LOF_A_14_30	2014, August	No	44	8.3
Lofoten	LOF_A_14_33	2014, August	No	46	8.8
Lofoten	LOF_A_14_41	2014, August	No	45	8.6
Lofoten	LOF_A_14_43	2014, August	No	195	37.0
North East Arctic	LOF_M_14_26	2014, March	Yes	46	8.7
North East Arctic	LOF_M_14_27	2014, March	Yes	47	8.9
North East Arctic	LOF_M_14_28	2014, March	Yes	44	8.3
North East Arctic	LOF_M_14_29	2014, March	Yes	46	8.6
North East Arctic	LOF_M_14_30	2014, March	Yes	43	8.1
North East Arctic	LOF_M_14_31	2014, March	Yes	50	9.4
North East Arctic	LOF_M_14_32	2014, March	Yes	50	9.4
North East Arctic	LOF_M_14_33	2014, March	Yes	50	9.4
North East Arctic	LOF_M_14_35	2014, March	Yes	42	8.0
North East Arctic	LOF_M_14_36	2014, March	Yes	53	10.0
North East Arctic	LOF_M_14_43	2014, March	Yes	88	16.7
North East Arctic	LOF_M_14_44	2014, March	Yes	47	9.0
North East Arctic	LOF_M_14_45	2014, March	Yes	45	8.6
North East Arctic	LOF_M_14_46	2014, March	Yes	57	10.7
North East Arctic	LOF_M_14_47	2014, March	Yes	50	9.4
North East Arctic	LOF_M_14_50	2014, March	Yes	39	7.4
North East Arctic	LOF_M_14_51	2014, March	Yes	43	8.1
North East Arctic	LOF_M_14_52	2014, March	Yes	44	8.3
North East Arctic	LOF_M_14_53	2014, March	Yes	24	4.5
North East Arctic	LOF_M_14_54	2014, March	Yes	43	8.2
North East Arctic	LOF_M_14_55	2014, March	Yes	46	8.7
North East Arctic	LOF_M_14_56	2014, March	Yes	55	10.5
North East Arctic	LOF_M_14_62	2014, March	Yes	50	9.4
North East Arctic	LOF_M_14_68	2014, March	Yes	38	7.2
Øresund	ORE_301	2012, March	Yes	44	8.2
Øresund	ORE_302	2012, March	Yes	42	7.7
Øresund	ORE_303	2012, March	Yes	40	7.6
Øresund	ORE_308	2012, March	Yes	33	6.0
Øresund	ORE_309	2012, March	Yes	45	8.4
Øresund	ORE_310	2012, March	Yes	39	7.4
Øresund	ORE_313	2012, March	Yes	105	19.8
Øresund	ORE_314	2012, March	Yes	34	6.0
Øresund	ORE_315	2012, March	Yes	25	4.7
Øresund	ORE_316	2012, March	Yes	49	9.3
Øresund	ORE_317	2012, March	Yes	38	7.1
Øresund	ORE_318	2012, March	Yes	36	6.6
Øresund	ORE_322	2012, March	Yes	33	6.0
Øresund	ORE_323	2012, March	Yes	52	9.7
Øresund	ORE_325	2012, March	Yes	46	8.6
Øresund	ORE_326	2012, March	Yes	39	7.2
Øresund	ORE_331	2012, March	Yes	50	9.3

Øresund	ORE_332	2012, March	Yes	49	9.2
Øresund	ORE_333	2012, March	Yes	48	8.9
Øresund	ORE_336	2012, March	Yes	41	7.7
Øresund	ORE_341	2012, March	Yes	45	8.4
North Sea	SOD_01	2002, March	Yes	43	8.3
North Sea	SOD_02	2002, March	Yes	46	8.8
North Sea	SOD_03	2002, March	Yes	44	8.3
North Sea	SOD_04	2002, March	Yes	55	10.4
North Sea	SOD_06	2002, March	Yes	45	8.7
North Sea	SOD_07	2002, March	Yes	46	8.8
North Sea	SOD_08	2002, March	Yes	42	8.1
North Sea	SOD_09	2002, March	Yes	49	9.4
North Sea	SOD_10	2002, March	Yes	42	8.1
North Sea	SOD_13	2002, March	Yes	43	8.3
North Sea	SOD_14	2002, March	Yes	48	9.2
North Sea	SOD_15	2002, March	Yes	45	8.5
North Sea	SOD_17	2002, March	Yes	46	8.7
North Sea	SOD_19	2002, March	Yes	47	8.8
North Sea	SOD_20	2002, March	Yes	39	7.2
North Sea	SOD_21	2002, March	Yes	45	8.3
North Sea	SOD_22	2002, March	Yes	46	8.8
North Sea	SOD_23	2002, March	Yes	44	8.4
North Sea	SOD_25	2002, March	Yes	45	8.6
North Sea	SOD_26	2002, March	Yes	53	10.0
North Sea	SOD_27	2002, March	Yes	51	9.0
North Sea	SOD_28	2002, March	Yes	46	8.6
North Sea	SOD_29	2002, March	Yes	46	8.8
North Sea	SOD_30	2002, March	Yes	42	7.6



## **DNA extraction and library creation of ancient Atlantic cod samples**

All extraction and library protocols were performed in a dedicated laboratory at the Department of Biosciences, University of Oslo following strict aDNA precautions (79, 80). This is a laboratory that is physically separated from the modern DNA laboratories and in which no modern samples have ever been processed. Extraction of ancient Atlantic cod bones ( $n = 19$ ) used a combined bleach and pre-digestion protocol (54). Each sample was exposed to UV for 10 minutes on each side, resulting in a total dosage of 4800 J/m<sup>2</sup> before being cut and milled to powder. Then, for each sample, two times 150-200 mg of bone powder (milled in a Retsch MM400) was incubated in 0.5% bleach solution for 15 min (94). The samples were subsequently washed with H<sub>2</sub>O and the remaining bone powder was exposed to a 30-minute pre-digestion treatment followed by an overnight, second digestion using a freshly prepared digestion buffer (95). Following the second digestion, the two eluates were combined and concentrated (Amicon-30kDA Centrifugal Filter Units) after which DNA was extracted using Qiagen Minelute columns according to manufacturer's instructions. DNA was eluted in 60 µl pre-heated (60°C) EB buffer with a 15 min incubation at 37°C. Negative controls were included in all extraction experiments. Blunt-end Illumina libraries were built (81), following (20) and ligated DNA was amplified using sample-specific seven base-pair indexes in the P7 primer to allow multiplexing. PCRs were done in 15 µl (2.5 U PfuTurbo Cx Hotstart DNA Polymerase (Agilent Technologies), 1x buffer, 0.2 mM per dNTP, 0.2 µM P7 index primer, 0.2 µM P5 IS4 primer and 0.4 mg/ml BSA) for 13 cycles (2 min at 95°C, 13 cycles of 30s at 95°C, 30s at 60°C and 70s at 72°C with a final extension of 10 min at 72°C). Amplified products were cleaned using Agencourt® AMPure XP beads at a 1:1.7 ratio, eluted in 30 µl in EB buffer and quantified using a Bioanalyzer 2100 (Agilent). Libraries were sequenced on an Illumina Hiseq 2500 at the Norwegian Sequencing Centre (125 bp paired-end) and demultiplexed allowing zero mismatches in the index tag.

## Figures and Tables

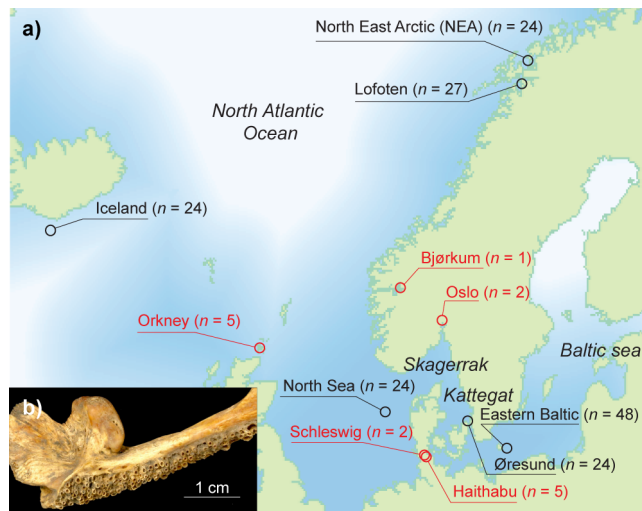


Figure 1.

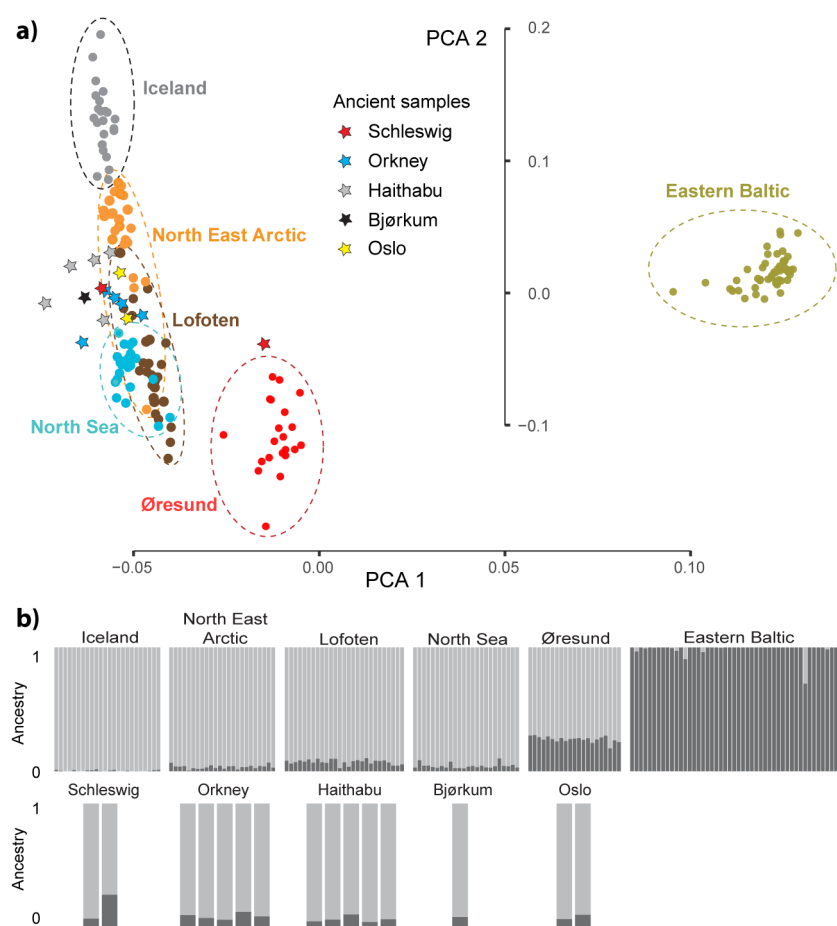


Figure 2.

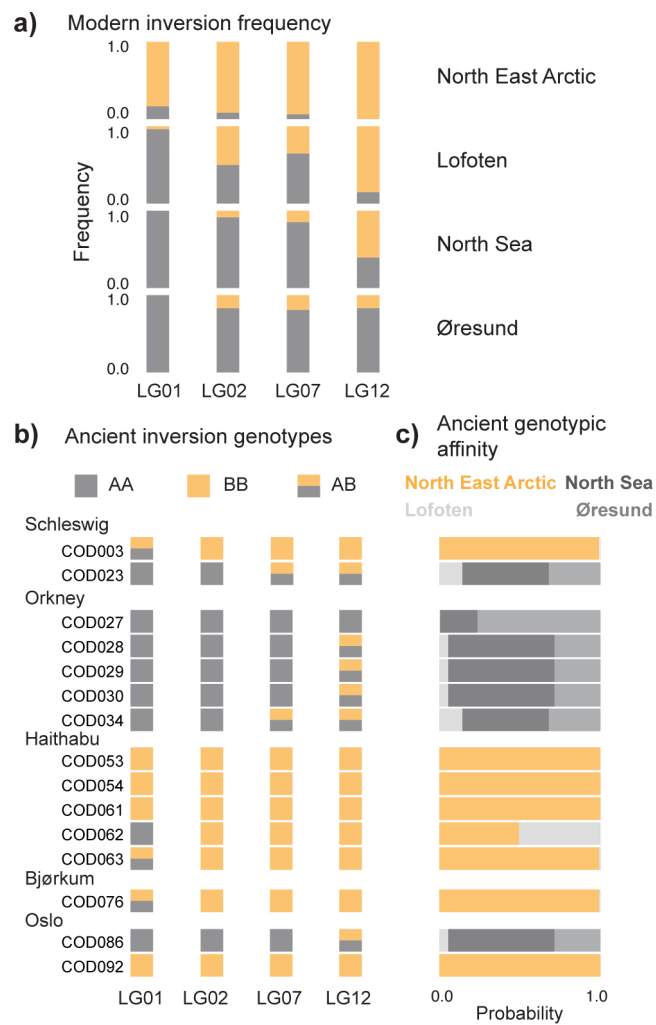


Figure 3.

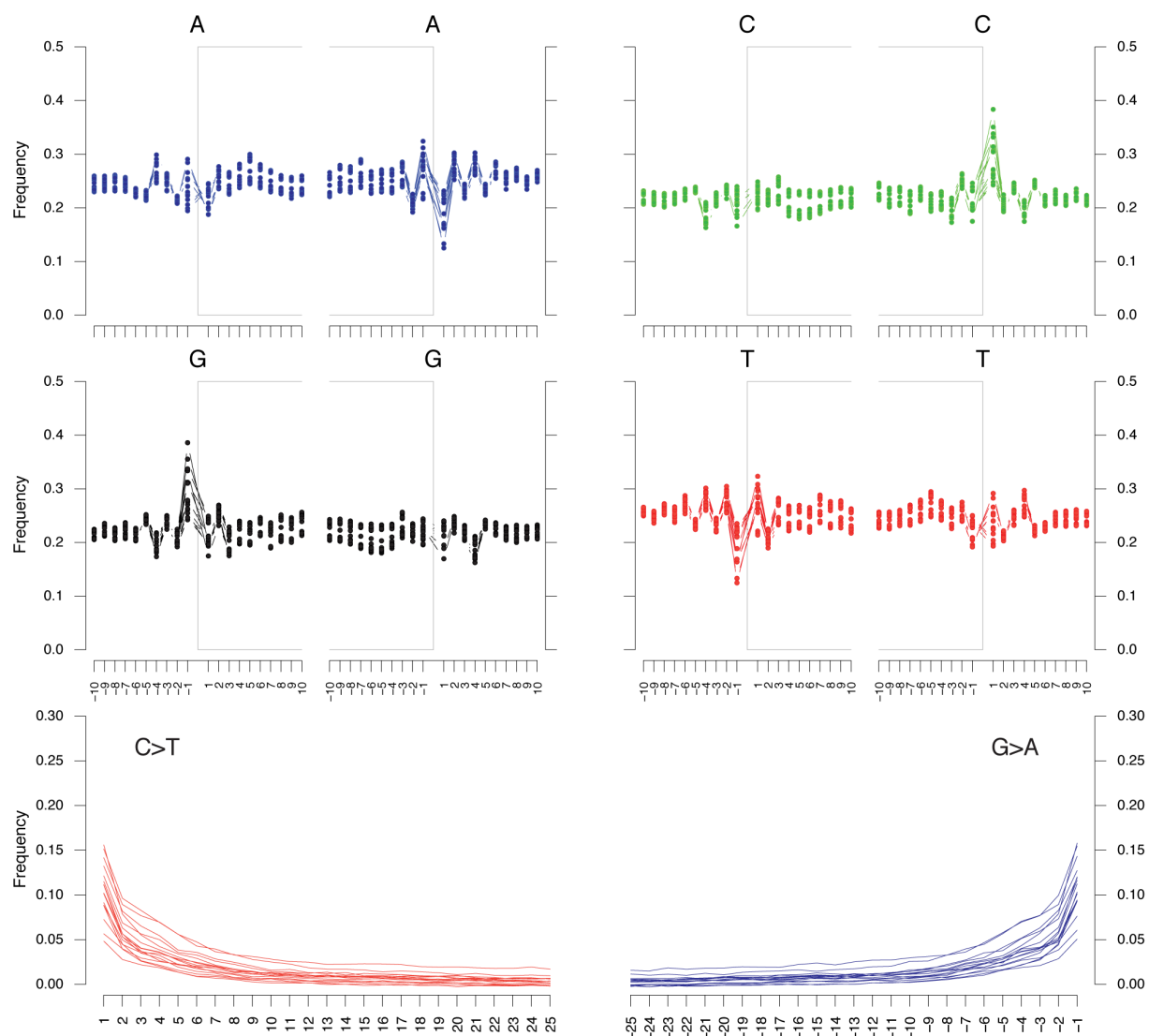


Figure S1.

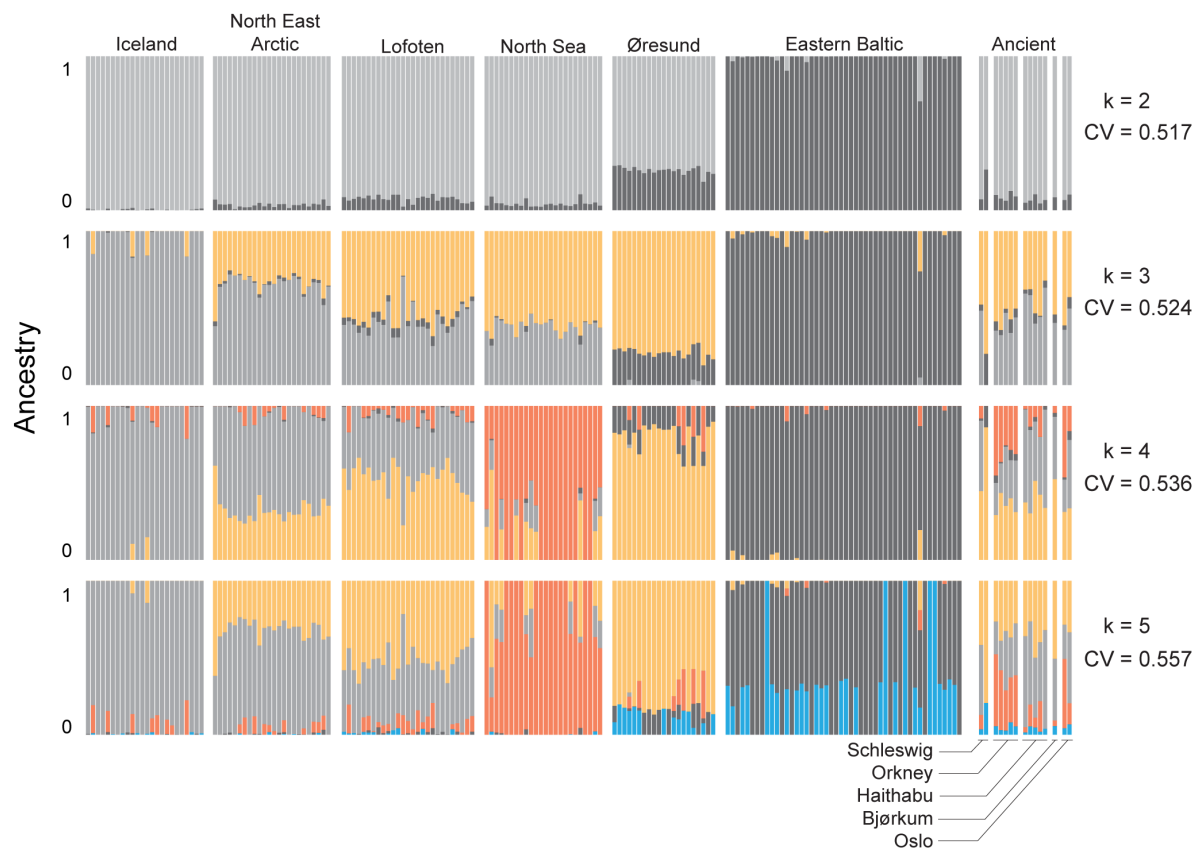


Figure S2.

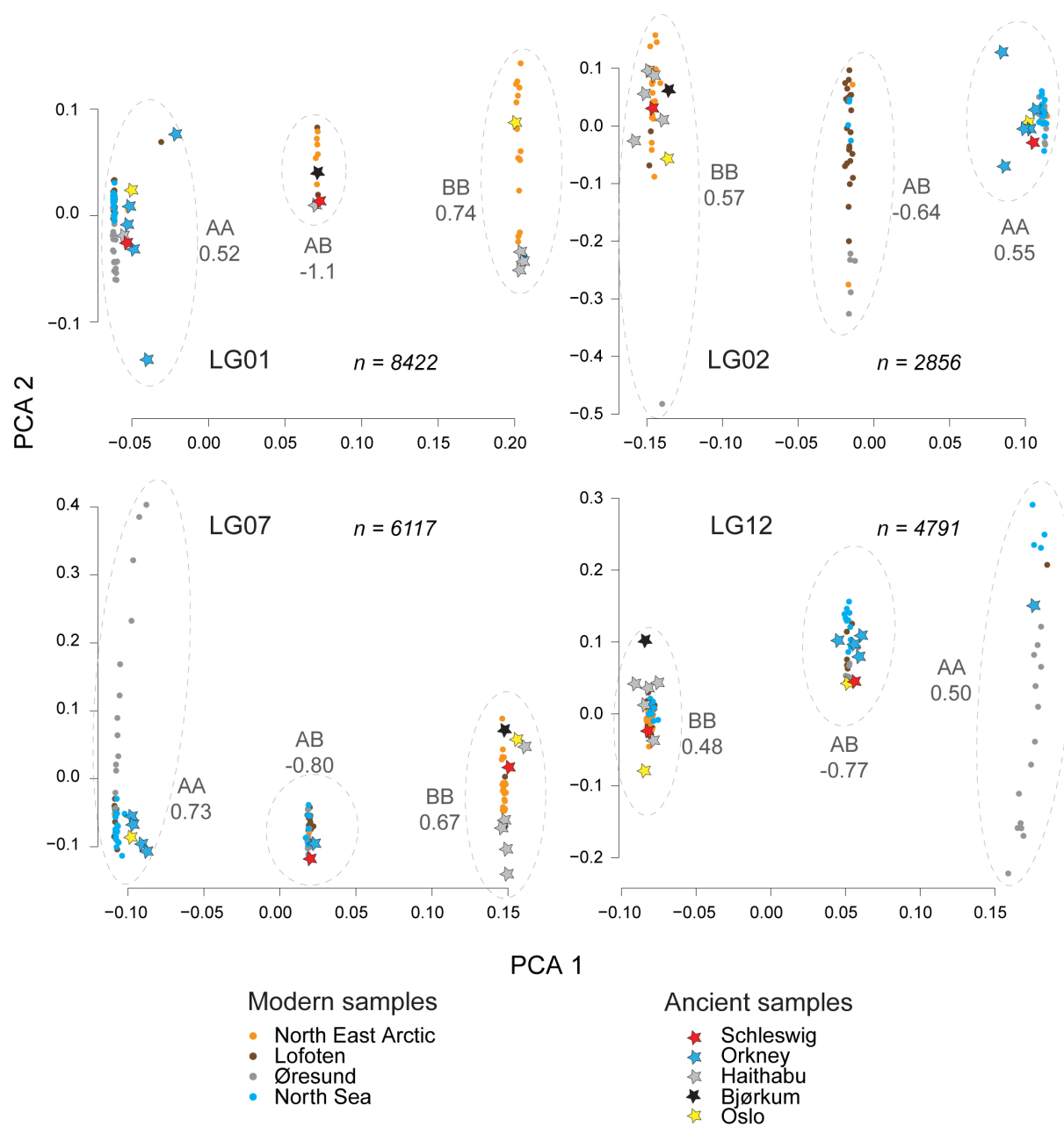


Figure S3.

**Table S1.** Specimen ID, location, estimated date and bone type of ancient Atlantic cod samples. All bones were morphologically identified as Atlantic cod (*Gadus morhua*). WGS shotgun libraries were paired-end sequenced, and we report the number of collapsed reads, their clonality, their endogenous DNA content (defined as the unique, non-repetitive fraction of reads aligning towards the gadmor2 reference genome with a minimum MapQ value of 25), the average insert length and the fold coverage obtained for the nuclear genome. Four other specimens (not shown) were extracted of which two did not yield libraries, and two had endogenous DNA content below 1%.

Specimen	Country	Location	Date (CE)	Bone type	Reads (millions)	Clonality (%)	Endogenous DNA (%)	Average insert length (bp)	Fold coverage
COD003	Germany	Schleswig	c.1100-1200	Vertebra	52	14	33	69	1.9
COD023	Germany	Schleswig	c.1200-1280	Articular	67	6	37	51	2.0
COD027	UK	Orkney	c.1000-1200	Premaxilla	59	9	42	75	2.9
COD028	UK	Orkney	c.1000-1200	Premaxilla	84	4	15	72	1.4
COD029	UK	Orkney	c.1000-1200	Premaxilla	58	5	33	71	2.1
COD030	UK	Orkney	c.1000-1200	Premaxilla	55	8	25	72	1.6
COD034	UK	Orkney	c.1000-1200	Premaxilla	49	23	27	72	1.5
COD053	Germany	Haithabu	c.800-1066	Cleithrum	83	8	28	47	1.7
COD054	Germany	Haithabu	c.800-1066	Cleithrum	58	8	37	67	2.3
COD061	Germany	Haithabu	c.800-1066	Vertebra	72	11	24	40	1.1
COD062	Germany	Haithabu	c.800-1066	Vertebra	76	8	25	46	1.4
COD063	Germany	Haithabu	c.800-1066	Articular	95	6	32	46	2.2
COD076	Norway	Bjorkum	c.700-950	Dentary	74	27	18	48	1.0
COD086	Norway	Oslo	c.1025-1175	Ceratohyal	59	12	46	78	3.4
COD092	Norway	Oslo	c.1025-1175	Cleithra	72	16	21	47	1.2



**Table S2.** Population, specimen ID and sample date of modern Atlantic cod specimens. We report if sampling took place during spawning, the number of reads obtained and the resulting fold coverage for the nuclear genome.

Population	Specimen ID	Sample Date	Spawning population	Number of reads (millions)	Fold coverage
Eastern Baltic	ARK_4001	2012, May	Yes	40	7.4
Eastern Baltic	ARK_4002	2012, May	Yes	20	3.8
Eastern Baltic	ARK_4003	2012, May	Yes	38	6.9
Eastern Baltic	ARK_4006	2012, May	Yes	34	6.3
Eastern Baltic	ARK_4007	2012, May	Yes	34	6.4
Eastern Baltic	ARK_4008	2012, May	Yes	47	8.8
Eastern Baltic	ARK_4010	2012, May	Yes	31	5.9
Eastern Baltic	ARK_4011	2012, May	Yes	49	9.1
Eastern Baltic	ARK_4012	2012, May	Yes	35	6.6
Eastern Baltic	ARK_4013	2012, May	Yes	37	6.9
Eastern Baltic	ARK_4014	2012, May	Yes	34	6.1
Eastern Baltic	ARK_4015	2012, May	Yes	44	8.3
Eastern Baltic	ARK_4016	2012, May	Yes	29	5.1
Eastern Baltic	ARK_4017	2012, May	Yes	45	8.5
Eastern Baltic	ARK_4018	2012, May	Yes	33	6.1
Eastern Baltic	ARK_4019	2012, May	Yes	47	8.8
Eastern Baltic	ARK_4020	2012, May	Yes	23	4.1
Eastern Baltic	ARK_4021	2012, May	Yes	57	10.6
Eastern Baltic	ARK_4022	2012, May	Yes	34	6.4
Eastern Baltic	ARK_4024	2012, May	Yes	31	5.8
Eastern Baltic	ARK_4030	2012, May	Yes	34	6.5
Eastern Baltic	ARK_4032	2012, May	Yes	91	17.2
Eastern Baltic	ARK_4039	2012, May	Yes	63	11.9
Eastern Baltic	ARK_4043	2012, May	Yes	68	12.8
Eastern Baltic	BOR_90E	2011, April	Yes	44	8.3
Eastern Baltic	BOR_91E	2011, April	Yes	47	9.0
Eastern Baltic	BOR_74E	2011, May	Yes	45	8.6
Eastern Baltic	BOR_79E	2011, May	Yes	50	9.6
Eastern Baltic	BOR_60E	2012, April	Yes	46	8.7
Eastern Baltic	BOR_611E	2012, May	Yes	45	8.5
Eastern Baltic	BOR_613E	2012, May	Yes	43	8.2
Eastern Baltic	BOR_614E	2012, May	Yes	42	7.9
Eastern Baltic	BOR_615E	2012, May	Yes	42	8.0
Eastern Baltic	BOR_617E	2012, May	Yes	41	7.8
Eastern Baltic	BOR_621E	2012, May	Yes	41	7.8
Eastern Baltic	BOR_624E	2012, May	Yes	42	8.1
Eastern Baltic	BOR_630E	2012, May	Yes	40	7.5
Eastern Baltic	BOR_655E	2012, May	Yes	73	13.8
Eastern Baltic	BOR_659E	2012, May	Yes	59	11.1
Eastern Baltic	BOR_664E	2012, May	Yes	66	12.5
Eastern Baltic	BOR_AL713	2012, May	Yes	44	8.4
Eastern Baltic	BOR_739E	2012, May	Yes	57	10.8
Eastern Baltic	BOR_749E	2012, May	Yes	59	11.2
Eastern Baltic	BOR_760E	2012, May	Yes	6	1.1
Eastern Baltic	BOR_AL724	2012, May	Yes	49	9.3
Eastern Baltic	BOR_AL736	2012, May	Yes	44	8.4
Eastern Baltic	BOR_AL741	2012, May	Yes	46	8.7
Eastern Baltic	BOR_AL777	2012, May	Yes	43	8.3
Iceland	I50_02	2003, April	Yes	49	10.9
Iceland	I50_03	2003, April	Yes	43	9.7
Iceland	I50_04	2003, April	Yes	44	9.8
Iceland	I50_05	2003, April	Yes	46	10.3
Iceland	I50_06	2003, April	Yes	32	7.1
Iceland	I50_07	2003, April	Yes	42	9.4
Iceland	I50_08	2003, April	Yes	35	7.7
Iceland	I50_09	2003, April	Yes	51	11.2
Iceland	I50_10	2003, April	Yes	43	9.6
Iceland	I50_11	2003, April	Yes	50	11.1
Iceland	I50_12	2003, April	Yes	39	8.8
Iceland	I50_13	2003, April	Yes	48	10.7
Iceland	I50_14	2003, April	Yes	26	5.7
Iceland	I50_15	2003, April	Yes	41	9.3
Iceland	I50_16	2003, April	Yes	49	11.0
Iceland	I50_17	2003, April	Yes	43	9.5
Iceland	I50_18	2003, April	Yes	46	10.1

Iceland	I50_19	2003, April	Yes	52	11.4
Iceland	I50_20	2003, April	Yes	54	12.1
Iceland	I50_23	2003, April	Yes	58	12.9
Iceland	I50_26	2003, April	Yes	53	10.3
Iceland	I50_38	2003, April	Yes	64	12.3
Iceland	I50_41	2003, April	Yes	92	19.1
Iceland	I50_42	2003, April	Yes	52	11.6
Lofoten	LOF_A_14_01	2014, August	No	60	11.4
Lofoten	LOF_A_14_03	2014, August	No	54	10.2
Lofoten	LOF_A_14_04	2014, August	No	47	9.0
Lofoten	LOF_A_14_05	2014, August	No	47	8.9
Lofoten	LOF_A_14_06	2014, August	No	52	10.0
Lofoten	LOF_A_14_08	2014, August	No	43	8.2
Lofoten	LOF_A_14_09	2014, August	No	50	9.5
Lofoten	LOF_A_14_10	2014, August	No	48	9.1
Lofoten	LOF_A_14_11	2014, August	No	53	10.1
Lofoten	LOF_A_14_16	2014, August	No	54	10.1
Lofoten	LOF_A_14_17	2014, August	No	60	11.4
Lofoten	LOF_A_14_18	2014, August	No	52	9.9
Lofoten	LOF_A_14_19	2014, August	No	54	10.3
Lofoten	LOF_A_14_20	2014, August	No	43	8.2
Lofoten	LOF_A_14_21	2014, August	No	52	9.8
Lofoten	LOF_A_14_22	2014, August	No	48	9.1
Lofoten	LOF_A_14_23	2014, August	No	46	8.8
Lofoten	LOF_A_14_24	2014, August	No	49	9.3
Lofoten	LOF_A_14_25	2014, August	No	59	11.2
Lofoten	LOF_A_14_26	2014, August	No	51	9.7
Lofoten	LOF_A_14_27	2014, August	No	65	12.4
Lofoten	LOF_A_14_28	2014, August	No	49	9.3
Lofoten	LOF_A_14_29	2014, August	No	46	8.7
Lofoten	LOF_A_14_30	2014, August	No	44	8.3
Lofoten	LOF_A_14_33	2014, August	No	46	8.8
Lofoten	LOF_A_14_41	2014, August	No	45	8.6
Lofoten	LOF_A_14_43	2014, August	No	195	37.0
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North East Arctic	LOF_M_14_27	2014, March	Yes	47	8.9
North East Arctic	LOF_M_14_28	2014, March	Yes	44	8.3
North East Arctic	LOF_M_14_29	2014, March	Yes	46	8.6
North East Arctic	LOF_M_14_30	2014, March	Yes	43	8.1
North East Arctic	LOF_M_14_31	2014, March	Yes	50	9.4
North East Arctic	LOF_M_14_32	2014, March	Yes	50	9.4
North East Arctic	LOF_M_14_33	2014, March	Yes	50	9.4
North East Arctic	LOF_M_14_35	2014, March	Yes	42	8.0
North East Arctic	LOF_M_14_36	2014, March	Yes	53	10.0
North East Arctic	LOF_M_14_43	2014, March	Yes	88	16.7
North East Arctic	LOF_M_14_44	2014, March	Yes	47	9.0
North East Arctic	LOF_M_14_45	2014, March	Yes	45	8.6
North East Arctic	LOF_M_14_46	2014, March	Yes	57	10.7
North East Arctic	LOF_M_14_47	2014, March	Yes	50	9.4
North East Arctic	LOF_M_14_50	2014, March	Yes	39	7.4
North East Arctic	LOF_M_14_51	2014, March	Yes	43	8.1
North East Arctic	LOF_M_14_52	2014, March	Yes	44	8.3
North East Arctic	LOF_M_14_53	2014, March	Yes	24	4.5
North East Arctic	LOF_M_14_54	2014, March	Yes	43	8.2
North East Arctic	LOF_M_14_55	2014, March	Yes	46	8.7
North East Arctic	LOF_M_14_56	2014, March	Yes	55	10.5
North East Arctic	LOF_M_14_62	2014, March	Yes	50	9.4
North East Arctic	LOF_M_14_68	2014, March	Yes	38	7.2
Øresund	ORE_301	2012, March	Yes	44	8.2
Øresund	ORE_302	2012, March	Yes	42	7.7
Øresund	ORE_303	2012, March	Yes	40	7.6
Øresund	ORE_308	2012, March	Yes	33	6.0
Øresund	ORE_309	2012, March	Yes	45	8.4
Øresund	ORE_310	2012, March	Yes	39	7.4
Øresund	ORE_313	2012, March	Yes	105	19.8
Øresund	ORE_314	2012, March	Yes	34	6.0
Øresund	ORE_315	2012, March	Yes	25	4.7
Øresund	ORE_316	2012, March	Yes	49	9.3
Øresund	ORE_317	2012, March	Yes	38	7.1
Øresund	ORE_318	2012, March	Yes	36	6.6
Øresund	ORE_322	2012, March	Yes	33	6.0
Øresund	ORE_323	2012, March	Yes	52	9.7
Øresund	ORE_325	2012, March	Yes	46	8.6
Øresund	ORE_326	2012, March	Yes	39	7.2
Øresund	ORE_331	2012, March	Yes	50	9.3
Øresund	ORE_332	2012, March	Yes	49	9.2

Øresund	ORE_333	2012, March	Yes	48	8.9
Øresund	ORE_336	2012, March	Yes	41	7.7
Øresund	ORE_341	2012, March	Yes	45	8.4
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North Sea	SOD_02	2002, March	Yes	46	8.8
North Sea	SOD_03	2002, March	Yes	44	8.3
North Sea	SOD_04	2002, March	Yes	55	10.4
North Sea	SOD_06	2002, March	Yes	45	8.7
North Sea	SOD_07	2002, March	Yes	46	8.8
North Sea	SOD_08	2002, March	Yes	42	8.1
North Sea	SOD_09	2002, March	Yes	49	9.4
North Sea	SOD_10	2002, March	Yes	42	8.1
North Sea	SOD_13	2002, March	Yes	43	8.3
North Sea	SOD_14	2002, March	Yes	48	9.2
North Sea	SOD_15	2002, March	Yes	45	8.5
North Sea	SOD_17	2002, March	Yes	46	8.7
North Sea	SOD_19	2002, March	Yes	47	8.8
North Sea	SOD_20	2002, March	Yes	39	7.2
North Sea	SOD_21	2002, March	Yes	45	8.3
North Sea	SOD_22	2002, March	Yes	46	8.8
North Sea	SOD_23	2002, March	Yes	44	8.4
North Sea	SOD_25	2002, March	Yes	45	8.6
North Sea	SOD_26	2002, March	Yes	53	10.0
North Sea	SOD_27	2002, March	Yes	51	9.0
North Sea	SOD_28	2002, March	Yes	46	8.6
North Sea	SOD_29	2002, March	Yes	46	8.8
North Sea	SOD_30	2002, March	Yes	42	7.6